INTRODUCTION

According to Ayurveda, medicinal plant products especially leaves, root, stem bark, flowers and seeds have been continuously used for the last so many years to treat number of animal and human diseases (Gupta et al., 2014; Maciel et al., 2002). Recently, number of medicinal plant products e.g. Azadirachta indica, Mimusops elengi, Calotropis gigantea etc. showed antimicrobial activity against specific bacterial/viral/fungal disease (Maithani et al., 2011; Gupta et al., 2014; Patil et al., 2014). To further explore the knowledge of medicinal plant products, researchers focused only on those active constituent extracted from various primary or secondary metabolites which is responsible for inducing antimicrobial/viral/bacterial activity against specific diseases (Gupta et al., 2014). These active metabolites could be isolated from the crude form of medicinal plant products and tested through various qualitative and quantitative based parameters i.e. TLC (thin layer chromatography); HPTLC (high performance thin layer chromatography) etc. Thus, the present work was totally focused on Acacia catechu with emphasis on its anti-viral properties in human peripheral blood mononuclear cells (PBMC).

One of poultry infectious diseases i.e. new castle disease virus (NDV, single-stranded RNA virus; family Paramyxoviridae) is reported worldwide with several consequences including an increase in mortality rates (Gupta et al., 2014). Recently, treatments are not available for new castle disease virus however, vaccination is an effective method to control this disease but it does not provide any hundred percent protections. Thus there is a need to search for those medicinal plant products which restricted the load of NDV disease in poultry animals.

Acacia catechu (khair, family Fabaceae), medicinal plant and is popularly known for its astringent and antioxidant activities. The extracts of Acacia catechu exhibit various immunopharmacological effects like antipyretic, antidiarrhoeal, hypoglycemic, hepatoprotective; antioxidant activities etc. (Lakshmi and Rajendran, 2012; Ray et al., 2006; Guleria et al., 2011). The current study was focused only on
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the immunopharmacological activity of *Acacia catechu* on human PBMC against NDV.

**MATERIALS AND METHODS**

**Collection and preparation of plant material**

Fresh leaves of *Acacia catechu* was collected in the morning from Vidya Pratishthan’s garden in the month of October 2015 in Baramati region, District Pune, Maharashtra, India. After collection of fresh plant leaves, first of all washed in running tap water in order to remove the dust particles and then with distilled water and finally dried in a shady area at room temperature. Afterwards, leaves were macerated with liquid nitrogen to prepare fine powder and were used for subsequent immunopharmacological assays.

**Qualitative and quantitative estimation of metabolites**

Secondary metabolites were determined qualitatively in the aqueous leaves extract of *Acacia catechu*. Different assays were performed qualitatively and revealed the presence of flavonoids (alkaline reagent test); terpenoids (acetic anhydride test) and phenolic compounds (ferric chloride test).

**Collection of NDV samples**

Poultry suspected disease samples of NDV were collected from suspected birds and these samples were studied under Biovillage programme scheme, Vidya Pratishthan’s School of Biotechnology, Baramati, India. For these studies, pathogen free chicken eggs were used and these eggs were purchased from Venkys India Ltd for segregation and propagation of NDV from field samples using allantoic cavity route of embryonated (9-11 day old) chicken eggs (Gupta *et al.*, 2014). These eggs were continuously observed in dark at regular time intervals, only bigger sized embryos were selected for inoculation. Embryo motility was observed continuously every 4h through candling after insertion of supernatant (200 µL) at 45° angle into embryonated chicken eggs. After the death of embryos, amnio-allantoic fluid was harvested and identified the presence of virus which is determined through haemagglutination (HA, 128) titre.

**Cytotoxicity and PBMC proliferation assay using NDV**

Anti-coagulant EDTA human blood samples were collected from Mangal Pathology Laboratory, Maharashtra, India and separated PBMC by means of density gradient centrifugation and then cultured for 48h in 96 well plates with variable doses of aqueous leaves extract (0.5-30mg/mL, 50µL) of *Acacia catechu* along with or without NDV (1:80 dilution, 10µL). Thereafter, centrifuging the plates (96 well) at 1800rpm for 8min at 4°C and add fresh complete medium into the 96-well plates. Again, incubating the plates for another 4h along with MTT (5mg/mL, 10µL). Afterwards, plates were centrifuged with discarding the supernatant, collecting the pellet and finally dispersing or dissolved in dimethyl sulfoxide (DMSO) solution. The optical density was measured at 570nm (Gupta and Chaphalkar, 2016; Gupta and Chaphalkar, 2015).

**CD14 monocyte surface marker**

Similarly, PBMC were cultured with variable concentration of aqueous leaves extract of *Acacia catechu* along with or without NDV (1:80 dilution, 10µL) for 48h in 96-well plates. After incubation, treated and non treated PBMC samples of *Acacia catechu* were collected and stained with CD14 FITC (3µL) monoclonal antibody. The samples were then incubated (at room temperature), lysed (red cell lysis buffer containing ammonium chloride, potassium bicarbonate and EDTA) and washed with PBS (pH, 7.2). The resulting stained cell pellet was resuspended in 500µL of PBS and run on a FACS Calibur flow cytometer (Gupta *et al.*, 2015).

**Statistical analysis**

All values were mentioned as Mean ± S.E. Data was represented by one-way ANOVA test (Boniferroni multiple comparison test).

**RESULTS AND DISCUSSION**

**Cytotoxicity and PBMC proliferation (NDV) assay**

The effect of variable doses of aqueous leaves extract of *Acacia catechu* on human PBMC proliferation assay with NDV and also determined its cytotoxicity in absence of NDV containing aqueous leaves extract on PBMC.
Cytotoxic and Antiviral Activity

using MTT (Figure 1). For cytotoxicity based studies, aqueous leaves extract showed a dose-dependent decrease in PBMC population at higher doses as compared to control where as PBMC cultured along with NDV containing variable concentration of aqueous leaves extract showed enhancement in proliferation at lower doses but there is significantly decline in NDV proliferation at higher doses as compared to control. In addition, NDV showed a significant enhancement of PBMC proliferation as compared to control. Overall, the data represents that Acacia catechu at higher doses showed cytotoxic as well as anti-viral activity at higher doses.

CD14 monocyte surface marker

The effect of aqueous leaves extract of CD14 monocyte surface marker on PBMC with or without NDV (Figure 2). There was a significant decrease in CD14 monocyte surface marker at higher doses of Acacia catechu against NDV as compared to control.

The present study describes about the importance of aqueous leaves extract of Acacia catechu and showed its safe and effective anti-viral agent against NDV. Normally, the virus cannot be able to mutate with the medicinal plants because of the complexity of chemicals e.g. Catechin, epicatechin, alkaloids, quercetin etc in the form of secondary metabolites that are present abundantly. As per the literature, medicinal plant i.e. Acacia catechu reported number of secondary metabolites e.g. terpenoids, flavonoids, phenolics etc that are present and showed its importance to provide potentially new chemotherapeutic agents against various dreadful pathogens.

According to Ayurveda, lot of research work has already been done related to immunopharmacological studies of medicinal plants with respect to humoral and cell mediated immune response using non specific antigen. These studies are of great importance for many immunopharmacologists pertaining to anti-viral, anti-inflammatory, anti-diabetic studies etc (Haider et al., 2011, Gupta et al., 2014). For these studies, our group used human PBMC (monocytes) which showed morphological heterogeneity, such as variability of size including granularity and nuclear morphology and determined its cytotoxicity and CD14 monocyte surface marker using with or without NDV. Firstly, cytotoxicity assay are essential for determining the responses of human PBMC in presence or absence of specific antigen i.e. NDV. It is already established that exposure of test candidates on human PBMC triggers a permanent growth arrest through the process of accelerated senescence in a fraction of the cell population (Gupta et al., 2015).

In short, loss of cellular viability measured through caliometric based assay i.e. MTT emerged as a favored method for assessment of cytotoxicity (Gupta et al., 2006).

One of the parameters interconnected with our immune system i.e. CD14 monocyte surface marker which is determined through flow cytometric analysis and provides reliable information whether aqueous extract of Acacia catechu showed stimulatory or suppressive effect. Normally, viral pathogens in human increased the number of monocytes count in the form of CD14 (receptor for lipopolysaccharide) surface marker. Finally, flow cytometric results suggested that production of CD14 surface marker may depend on cell types (monocytes or macrophages) and their species origin, different cells having obviously individual necessity for signal transduction pathways. In particular, determination of CD14 marker from human PBMC assumed to play a considerable role in the pathophysiology of hormonal immune system. The findings of these preliminary studies showed that the aqueous extract of Acacia catechu against NDV virus and the results of anti-viral activity including its cytotoxic effect against NDV are shown in respective figures. The results indicate that aqueous leaves extract of Acacia catechu shows a dosage-dependent correlation and found that aqueous leaves extract could significantly reduce the CD14 count in human PBMC exposed to NDV. The results of our immunopharmacological studies on human PBMC after exposing with NDV suggest that the aqueous leaves extract shows anti-viral effect on human PBMC.

These studies suggest that aqueous leaves extract of Acacia catechu significantly inhibits proliferation assay and CD14 monocyte surface marker when using NDV.
Figure 1. Effect of variable doses of aqueous leaves extract of *Acacia catechu* on NDV in human PBMC. PBMC were cultured with or without optimized dose of NDV along with variable doses of aqueous root extract (0.5, 1, 10 and 30mg/mL, 50µL) or NDV alone. After 3 days, proliferation was measured by MTT assay. The results are presented as Mean ± S.E. *P < 0.05, **P < 0.01, ***P < 0.001 as compared to control.

Figure 2. Flow cytometric analysis of aqueous leaves extract extracted from *Acacia catechu* on CD14 monocyte surface marker. PBMC were treated with variable doses of aqueous leaves extract (0.5, 1, 10 and 30mg/mL, 50µL) along with or without NDV and then lysed and washed the cells with phosphate buffered saline and analyzed through flow cytometer (FACS, Calibur) using forward and side scatter gating applied for data acquisition of 10000 events of cell populations representing different phenotypes analyzed using cell quest software.
Further investigations of the aqueous leaves extract should be done through in vivo assessment for immunopharmacological (anti-viral) studies in mice models with identification of the major active candidates responsible for anti-microbial activities.

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